

# Rotavirus potency assay by CCID<sub>50</sub> method

WHO/PAHO workshop – June 2006

Geneviève Waeterloos, Scientific Collaborator  
Biological Standardisation  
Scientific Institute of Public Health  
Federal Public Service: Health, Food Chain Security and Environment  
Brussels - Belgium

# Rotavirus assay (CCID<sub>50</sub>)

- Objectives
- Titration method
- Validation data
- Batch release testing
- Additional comments



# Rotavirus assay (CCID<sub>50</sub>)

- Objectives
- Titration method
- Validation data
- Batch release testing
- Additional comments



# Rotavirus assay ( $CCID_{50}$ )

## Objectives

1. To determine the viral titre in rotavirus in the batch submitted for lot release
2. To determine the identity G1 of the vaccine strain



# Rotavirus assay (CCID<sub>50</sub>)

- Objectives
- Titration method
- Validation data
- Batch release testing
- Additional comments



# Rotavirus assay ( $CCID_{50}$ )

## ➤ Titration method principle

To determine the potency of Rotavirus by assessing the dose infecting 50% of MA-104 cell monolayer (monkey kidney -by end-point dilution) followed by revelation with MoAb 2C9 (anti-VP7) and immunoperoxidase staining or indirect immuno-fluorescence



# Rotavirus assay ( $CCID_{50}$ )

1. Preparation of cell substrate
2. Virus activation step (30min) dil 1/10  
& Successive dilutions in activation medium
3. Inoculation virus suspension & incubation at  $37 \pm 1^\circ C$  for  $7 \pm 1$  days



# Rotavirus assay ( $CCID_{50}$ )

4. Revelation: MoAb + immunoperoxidase or indirect immunofluorescence staining
5. Reading & Calculation of titres by Spearman-Kärber method (or Reed-Muench)( $\log CCID_{50}/\text{ml}$ )
6. Validity of the assay (criteria)
7. Acceptance of the batch (criteria)



# 1. Preparation of cell substrate

- MA-104 foetal monkey kidney cells
- cell growth medium:  
EMEM - 1% L-Glu - 10% FCS
- $50.000 \pm 10.000$  cells/ml
- 200 µl/well (microplates 96-wells)
- incubation 4 days at  $37 \pm 1C^\circ$  -  $5 \pm 1\%$   $CO_2$
- monolayer confluent (100%)



## 2. Virus activation step & dilutions

- Reconstitution of vaccines (3 vials + 3 "stability" and reference preparation (1 vial in triplicate) with 1ml WFI)
- dilution 1/10 in activation medium  
EMEM - 1% L-Glu -1% Ab - 0,32% trypsine
- 20-30 min at RT
- successive dilutions (1/10 - 1/4) - same medium



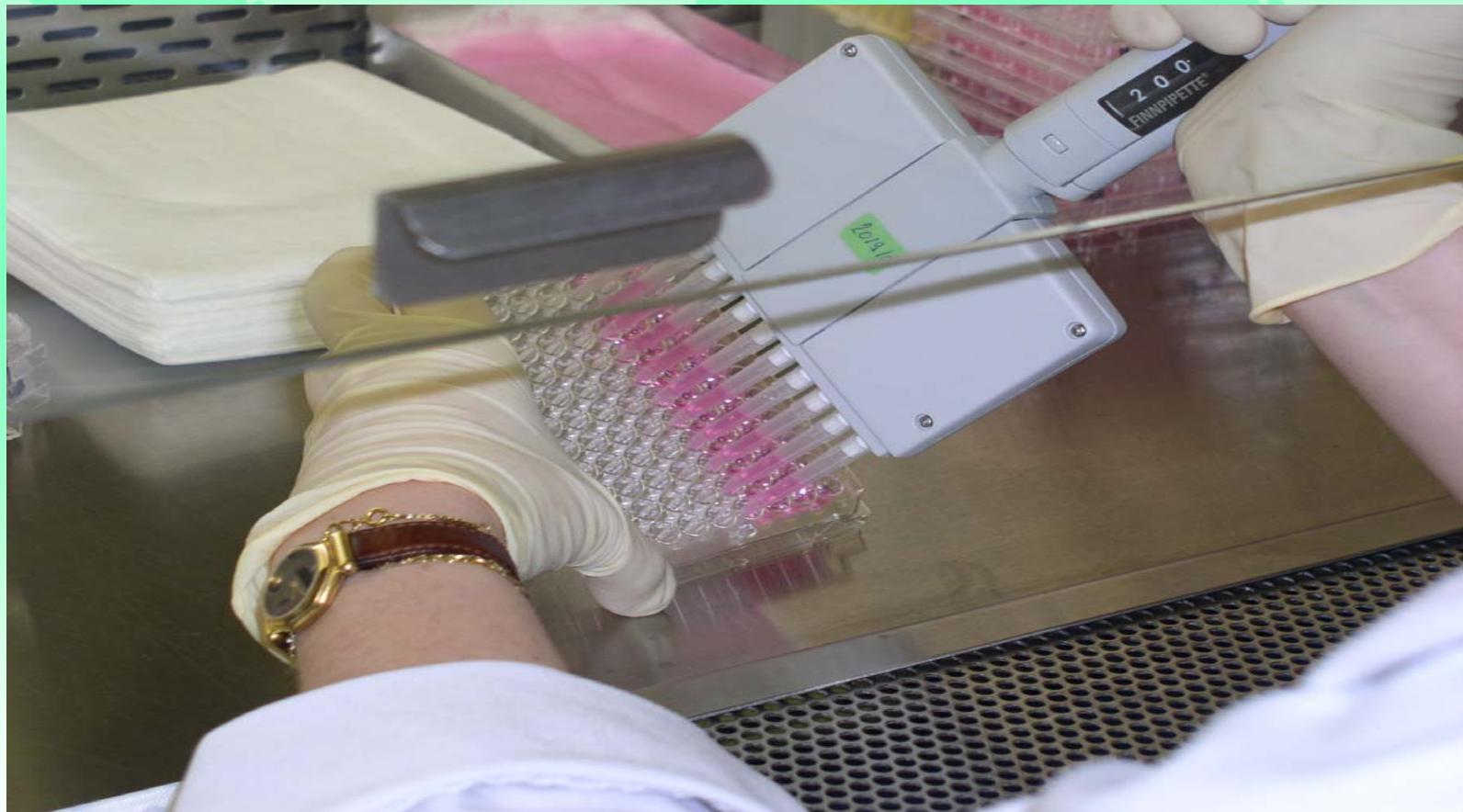
### 3. Inoculation of virus suspensions

- Microplates washed 2 times (150 µl/well)
- Carefully - multichannels
- Inoculation : 100µl/well - 10 wells/dilution -  
x dilutions/vial

| 1   | 2   | 3   | 4   | 5   | 6   | 7   | 8   | 9   | 10  | 11 | 12 |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|----|----|
| 4,6 | 4,6 | 4,6 | 4,6 | 4,6 | 4,6 | 4,6 | 4,6 | 4,6 | 4,6 |    | T  |
| 5,2 | 5,2 | 5,2 | 5,2 | 5,2 | 5,2 | 5,2 | 5,2 | 5,2 | 5,2 |    | T  |
| 5,8 | 5,8 | 5,8 | 5,8 | 5,8 | 5,8 | 5,8 | 5,8 | 5,8 | 5,8 |    | T  |
| 6,4 | 6,4 | 6,4 | 6,4 | 6,4 | 6,4 | 6,4 | 6,4 | 6,4 | 6,4 |    | T  |

- Incubation 7 days -  $37 \pm 1^\circ\text{C}$  -  $5 \pm 1\% \text{ CO}_2$  - HR

# Washing before virus inoculation



## 4. Revelation

- To wash microplates (150 µl/well)
- Fixation : iced acetone solution (80%) - 20 min at -20°C - 150µl/well
- Acetone discarded (kleenex) and let dry
- 100µl MoAb 2C9/well (1/250 in PBS w/o Ca<sup>++</sup> and Mg<sup>++</sup> with 5% skimmed milk = "blotto")
- Incubation 1 hour at 37± 1°C
- To wash plates 4 times with PBS w/o Ca<sup>++</sup> and Mg<sup>++</sup>

# Distribution of MoAb anti-VP7



## 4a. Revelation by immunoperoxidase

- Ab Anti-mouse IgG conjugated to peroxidase (1/400 in blotto) - 50µl/well - incubation 1 h - 37°C ± 1°C.
- To empty µplates & wash 4 times /PBSw/o
- Fresh solution : 1 tablet DAB + 1 tablet urea in 30 ml distilled water
- 50 µl/well incubation 10 min at RT (25± 5°C).
- To wash µpls 3 times in tap water
- To let dry (kleenex wypall)



## 4b. Revelation by indirect immunofluorescence staining

- anti-mouse IgG conjugated to FITC (1/200 in blotto + Evans Blue 1/300)
- 100µl/well - Incubation 1 hour at  $37\pm 1^\circ C$
- To wash µpls 4 times in tap water
- To let dry (kleenex wypall)

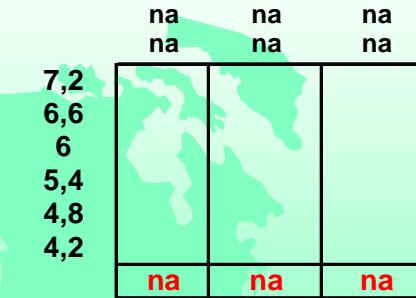


## 5. Reading & calculations

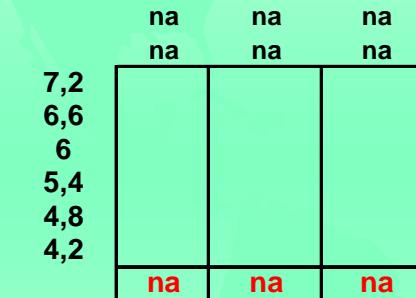
- reading : wells with stained cells = rotavirus positive
- calculation of titre by Reed-Muench or Spearman-Kärber
- estimation of the precision by Irwin-Cheesman



VACCIN  
**Frais**

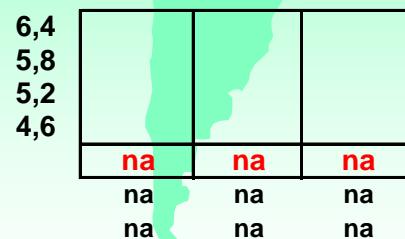


37°C



REFERENCE

Gsk  
RVC021A44/Q



Date du test:

-2004

Spécification : ? 6,2 logCCID<sub>50</sub>/1,0 ml

**Frais**

Titre moyen: => #DIV/0!  
95%: => #DIV/0!  
Limites: => #DIV/0! - #DIV/0!

**Stabilité**

Titre moyen: => #DIV/0!  
95%: => #DIV/0!  
Limites: => #DIV/0! - #DIV/0!

Perte de titre : => #DIV/0!  
Spécification: ? 0,5 logCCID<sub>50</sub>/1,0 ml

Gsk Titre : 6,5 logCCID<sub>50</sub>/ml

Titre moyen: => #####  
95%: => #####  
Limites: => ##### - #####

n\* = 0

# Spearman-Kärber

$$-\log CCID_{50} = -\log_{10}(d_1) - [\log_{10}(d) \times (Sp-0,5)]$$

Where

$d_1$  = highest dilution with 100% CPE

$d$  = dilution step (i.e. 0,6log)

$Sp$  = sum of proportion of all positive wells



# Spearmann-Kärber

| Nº lot | Dilution | Positive recorded |
|--------|----------|-------------------|
|        | -4,6     | 10                |
|        | -5,2     | 8                 |
|        | -5,8     | 2                 |
|        | -6,4     | 0                 |

$$-\log \text{CCID}_{50} = -4,6 - [0,6 \times ((10 + 8 + 2 + 0) / 10 - 0,5)]$$

$$-\log \text{CCID}_{50} = -4,6 - 0,9$$

$$-\log \text{CCID}_{50} = -5,5$$

$$+\log \text{CCID}_{50} = +5,5$$

Correction factor:  $\log_{10} 1000 \mu\text{l} / 100 \mu\text{l} = 1,0$

Vaccine titre :  $5,5 + 1,0 = 6,5 \log \text{CCID}_{50}/\text{ml}$

# Precision by Irwin-Cheesman

$$s = \log d \times \sqrt{ \sum [(p \times (1-p)) / n - 1] }$$

s = std deviation

d = dilution step

$\sum$  = sum of all dilutions

p = proportion of infected (positive) wells for one specific dilution

1 - p = proportion of non-infected (negative) wells

n = nr of inoculated well/dilution

# Precision by Irwin-Cheesman

| Dilution | Positive recorded | p     | 1-p   | p x (1-p) |
|----------|-------------------|-------|-------|-----------|
| -4,6     | 10                | 10/10 | 0/10  | 0         |
| -5,2     | 8                 | 8/10  | 2/10  | 16/100    |
| -5,8     | 2                 | 2/10  | 8/10  | 16/100    |
| -6,4     | 0                 | 0/10  | 10/10 | 0         |

$$\begin{aligned}s &= 0,6 \times \sqrt{(32/100) / 9} \\&= 0,6 \times 0,1886 \\&= 0,1132\end{aligned}$$

Confidence interval ( $P=0,95$ ) :  $0,1132 \times 1,96 = 0,22 = 0,2$

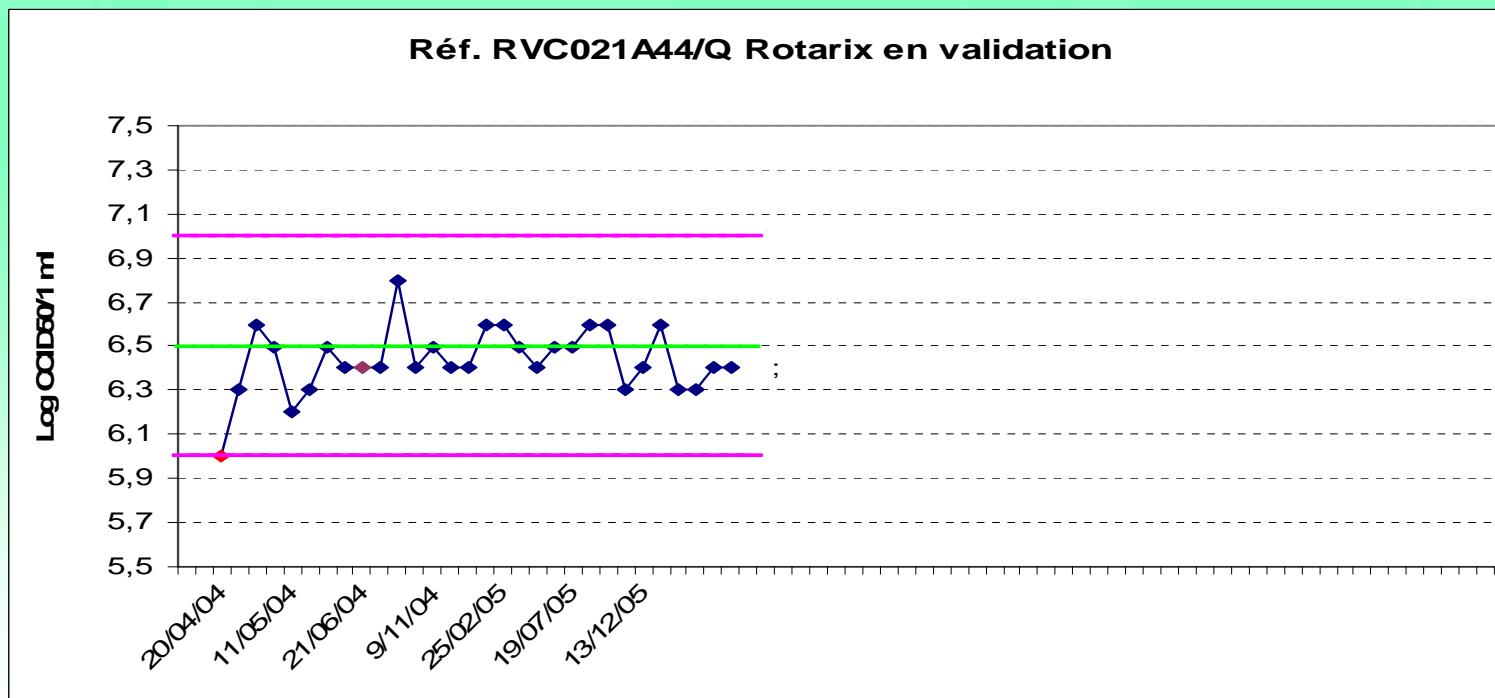
Virus titre :  $6,5 \pm 0,2 \log CCID_{50}/ dose$

## 6. Validity of the assay

- negative control cells : free of toxicity or abnormalities and do not show any fluorescent/coloured cells
- Homogeneous distribution of CPE vs dilutions (10-90 % of CPE on 3 dilutions)
- the confidence interval ( $P=0,95$ ) of the vaccine virus titre is  $\leq 0,3\log$  (\*)
- max diff.  $0,5\log$  between 2 vials/3 (\*)



➤ reference titre: Observed reference titre within 0,5 log of its established mean titre - Company:  $6,5 \pm 0,5$  logCCID50/ml or IPH titre: 6.4/1ml - LCL: 6.0/1ml - UCL: 6.9/1ml - n=30



## 7. Acceptance of the batch (criteria)

- the estimated titre of the batch meets the release specification :  
 $\geq 6,2 \log CCID_{50}/\text{ml}$
- the maximum loss of titre is  
 $0,5 \log CCID_{50}/\text{ml}$



# Rotavirus assay (CCID<sub>50</sub>)

- Objectives
- Titration method
- Validation data
- Batch release testing
- Additional comments



# Rotavirus assay ( $CCID_{50}$ )



## Validation data

1. The test specificity
2. The range of titration
3. The limit of detection
4. The precision (intra-assay repeatability)
5. The reproducibility (inter-assay repeatability)



# Rotavirus assay ( $CCID_{50}$ )

## 1. The test specificity

- MA-104 cells
- Trypsin activation step
- MoAb anti-VP7 & Validation data provided by Manufacturer (each new MoAb batch)
- File: demonstrated in identity test performed by seroneutralisation

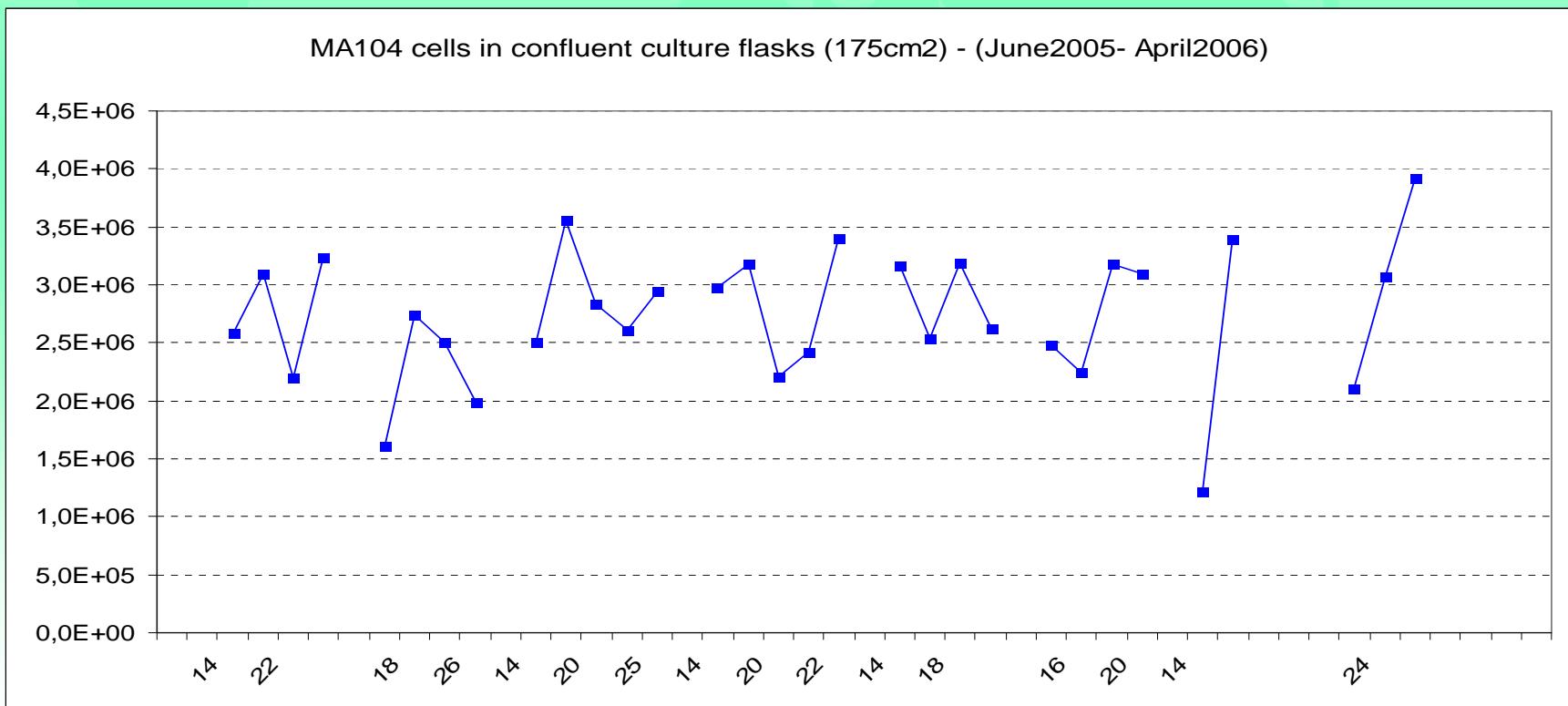


# Rotavirus assay ( $CCID_{50}$ )

Validation on MA-104 cells

cell bank (nr of passages)

cell concentration in culture flasks

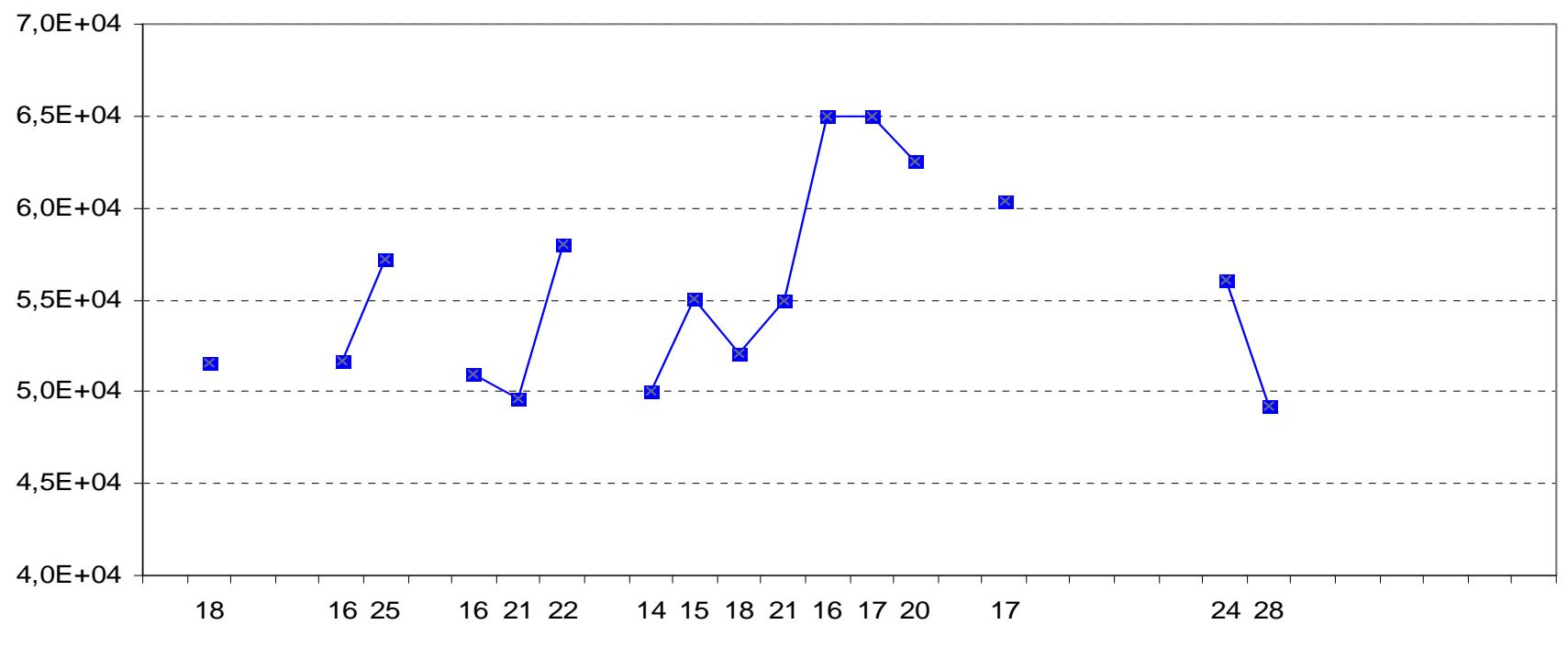


# Rotavirus assay (CCID<sub>50</sub>)

MA-104 cells

cell concentration inoculated  
(target 50.000+/-10.000)

MA104 cell concentration (50.000 +/- 10.000 cell/ml)



# Rotavirus assay ( $CCID_{50}$ )

## 2. The range of titration

In practice: choice of range of dilution

## 3. The limit of detection

End-point dilution method (0-100% CPE)



# Rotavirus assay (CCID<sub>50</sub>)

## 4. The precision (intra-assay repeatability)

one sample - x replicates - same day

| Date       | replicate 1 | replicate 2 | replicate 3 | mean | max diff | stdev |
|------------|-------------|-------------|-------------|------|----------|-------|
| 14/03/2006 | 6,2         | 6,5         | 6,3         | 6,3  | 0,3      | 0,15  |
| 28/03/2006 | 6,2         | 6,3         | 6,3         | 6,3  | 0,1      | 0,06  |
| 4/04/2006  | 6,6         | 6,6         | 6,7         | 6,6  | 0,1      | 0,06  |
| Mean       |             | 6,4         |             | 0,2  |          | 0,09  |

# Rotavirus assay ( $CCID_{50}$ )

## 5. The reproducibility (inter-assay repeatability) - time - operators

| RVC021A44/Q in-house ref.         |             |             |
|-----------------------------------|-------------|-------------|
| Nr                                | 30          |             |
| Mean                              | 6,4         |             |
| Stdev                             | 0,2         |             |
| cv (%):                           | 2,4%        |             |
| min                               | 6,0         |             |
| max                               | 6,8         |             |
|                                   | Lower Limit | Upper limit |
| $m \pm 2s = \text{alert limits}$  | 6,1         | 6,7         |
| $m \pm 3s = \text{action limits}$ | 6,0         | 6,9         |

# Rotavirus assay (CCID<sub>50</sub>)

- Objectives
- Titration method
- Validation data
- Batch release testing
- Additional comments



# Rotavirus batch release testing

1. Appearance
2. pH
3. Identity (by titration)
4. Potency
5. Thermal stability (7d -  $37\pm1^\circ\text{C}$ )

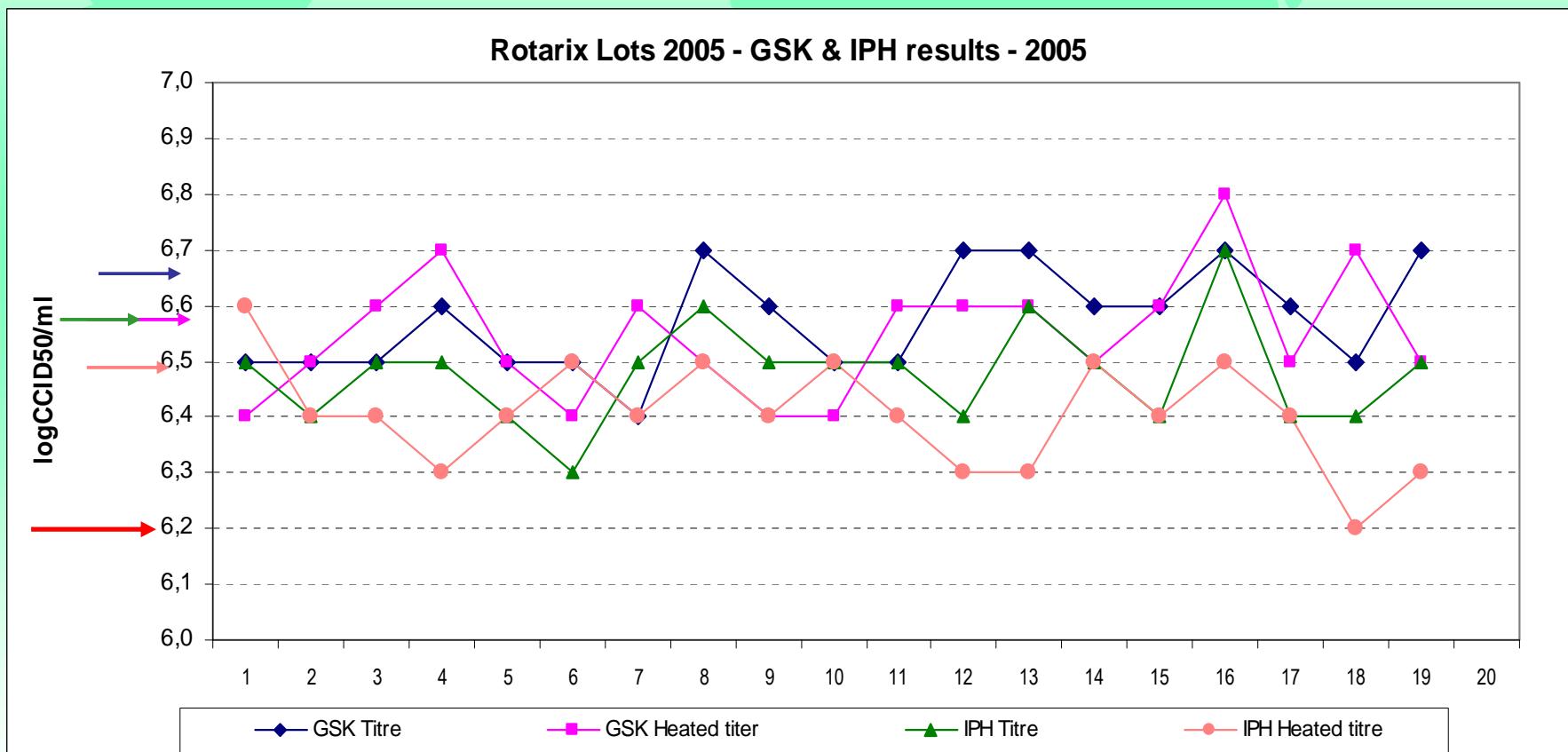


# Rotavirus batch release testing

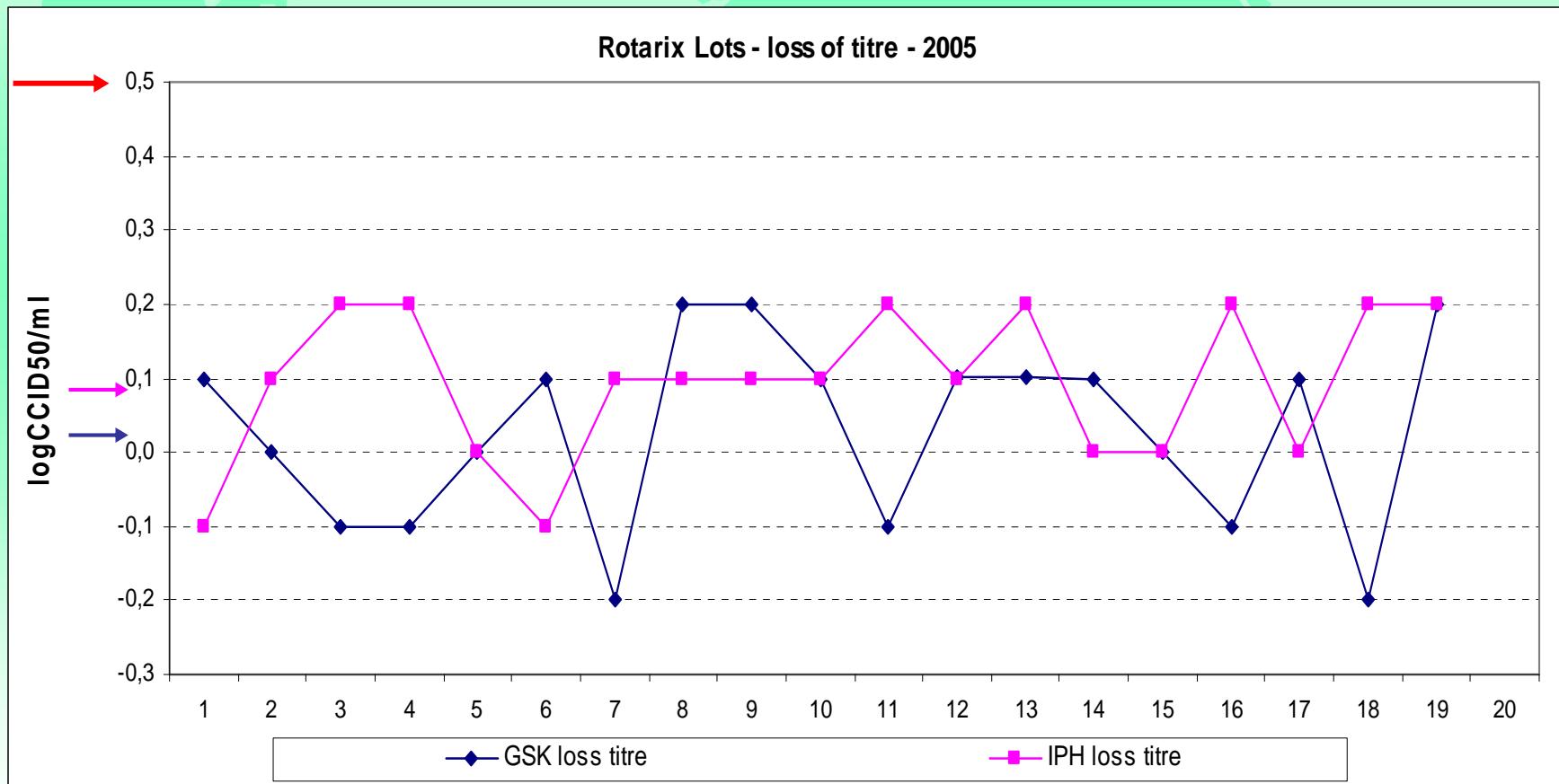
| Test                             | Specification   |
|----------------------------------|---|
| Appearance before reconstitution | Whitish cake or powder  |
| Appearance after reconstitution  | Clear colourless solution   |
| pH after reconstitution with WFI | Between 7,5 and 8,5   |
| Identity                         | Positive by titration   |
| Titre                            | $\geq 6,2 \log_{10} \text{CCID}_{50}/\text{ml}$                                   |
| Heated titre (7d, 37°C)          | To be determined ( $\log_{10} \text{CCID}_{50}/\text{ml}$ )                       |
| Titre loss                       | Not more than $0.5 \log_{10} \text{CCID}_{50}/\text{vial}$ from the release titre |



# Rotavirus batch release testing



# Rotavirus batch release testing



# Rotavirus assay (CCID<sub>50</sub>)

- Objectives
- Titration method
- Validation data
- Batch release testing
- Additional comments



# Additional comments

- Availability of qualified
  - ✓ MA-104 cell bank
  - ✓ Stock of MoAb 2C9
- Expensive equipment (Epi-fluorescence)
- Questions ?



# Rotavirus assay (CCID<sub>50</sub>)

MUCHAS GRACIAS POR  
SU ATTENCION

