



Rijksinstituut voor Volksgezondheid en Milieu Ministerie van Volksgezondheid, Welzijn en Sport

Interpretation of AMR surveillance data: sources of error and bias

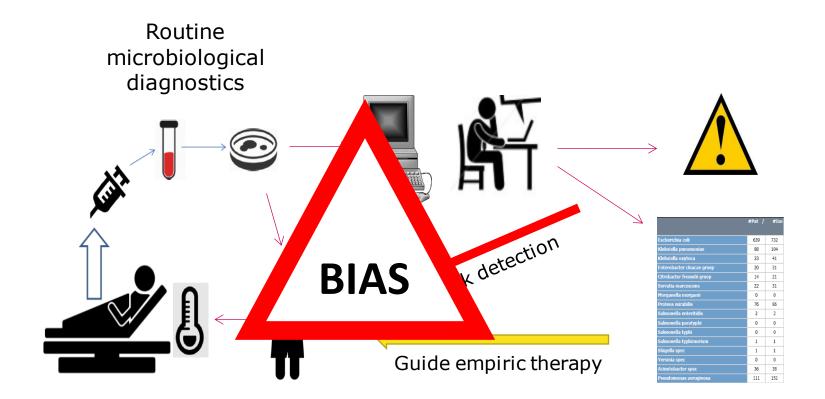
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AMR surveillance based on *routine diagnostics*





Surveillance using routine data

Advantages

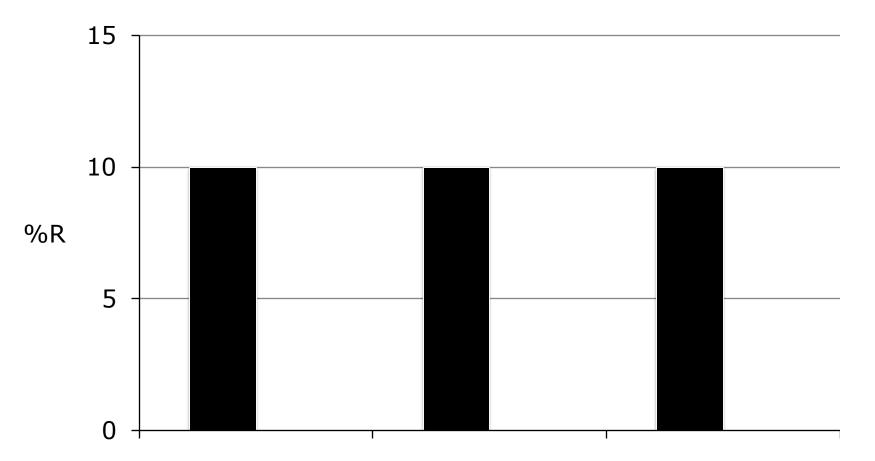
- > Large numbers for relatively little costs
- > Relatively insensitive to outbreaks
- Gives a good overall picture of resistance
- > Can be used to follow trends and emerging resistance
- > Guide antibiotic therapy and support infection prevention (under certain conditions!)

Disadvantages

- > No data on colonisation
- > Difficult to standardize
- > Risk of error and bias

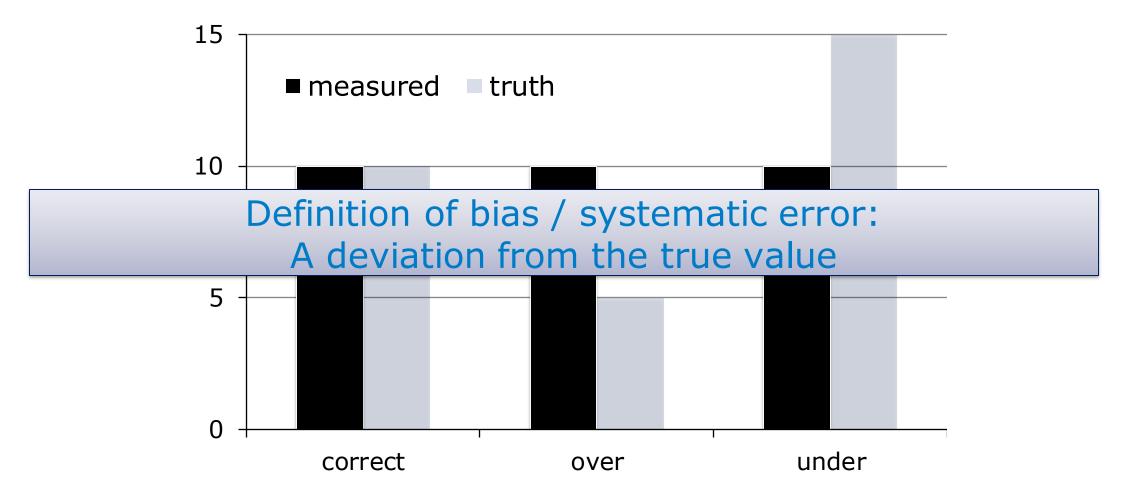


Is our estimate the truth?





Is our estimate the truth?





Representativeness



Type of hospital



Geographical region



Type of department



Season



Error and bias in national AMR surveillance

Surveillance system

- 1. Geographical region
- 2. Hospital types

Sampling procedures

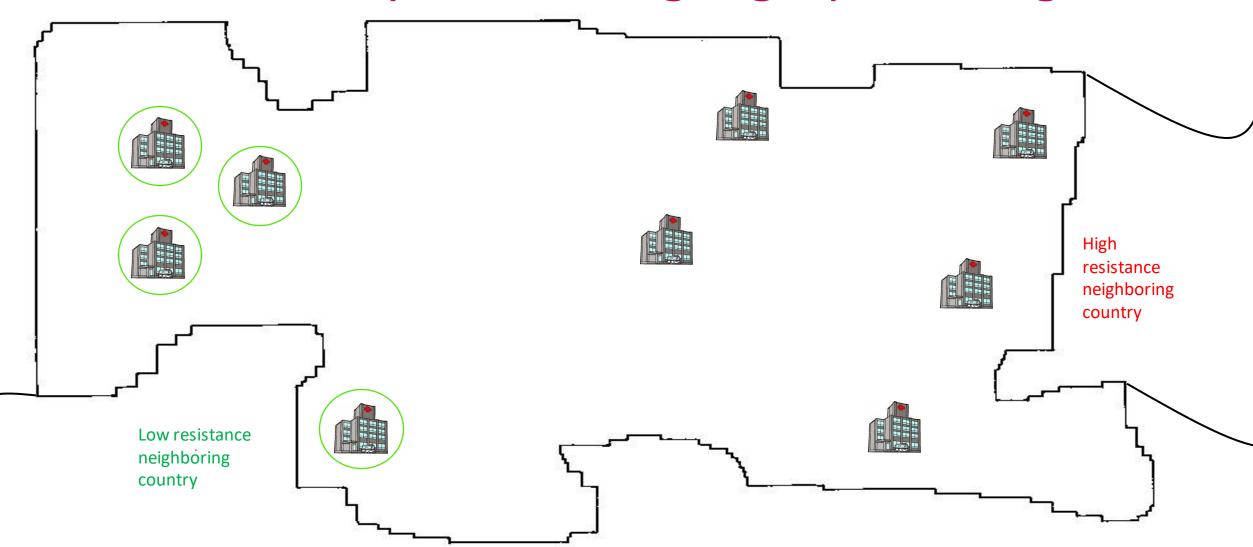
- 1. Selection of patients
- 2. Sampling after treatment
- 3. Follow-up samples
- 4. Sample size

Laboratory procedures

- 1. AST guidelines
- 2. Measurement error
- 3. Selective testing

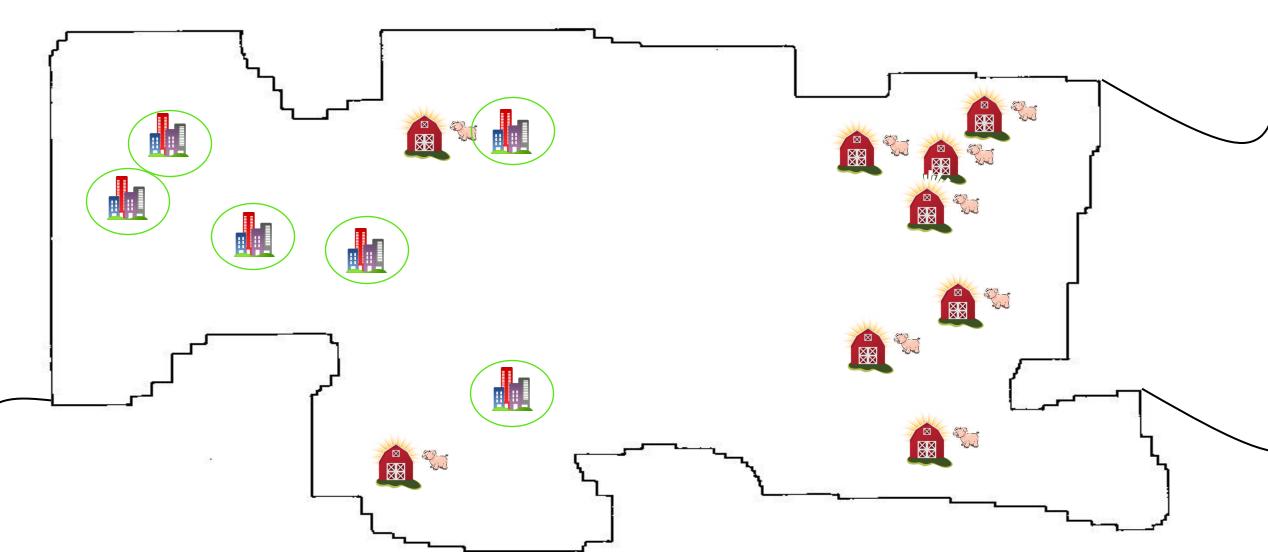


Surveillance system – 1. geographical region



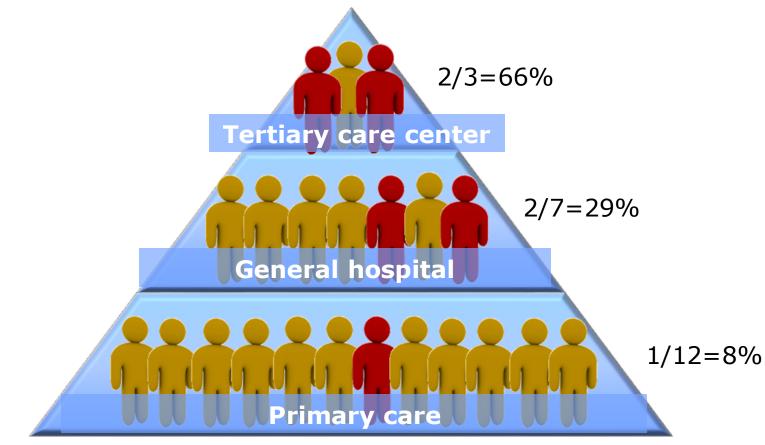


Surveillance system – 1. geographical region





Surveillance system – 2. hospital types



Overall resistance: 5/22 = 23%



Error and bias in national AMR surveillance

Surveillance system

- 1. Geographical region
- 2. Hospital types

Sampling procedures

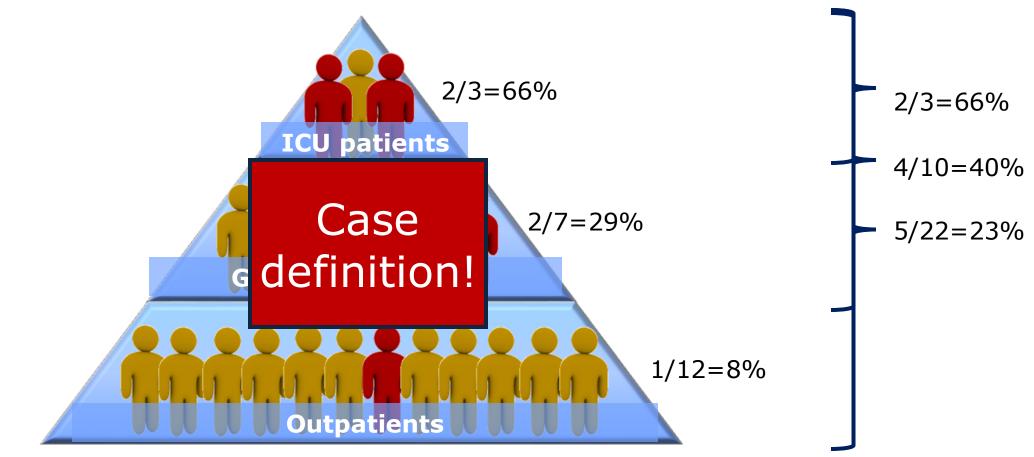
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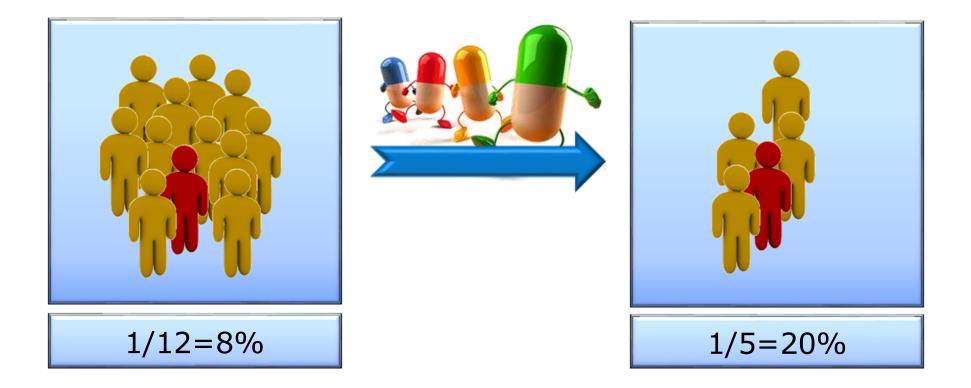
Sampling procedures – 1. patient population



Overall resistance: 5/22 = 23%



Sampling procedures – 2. sampling after treatment





Sampling procedures – 3. follow-up samples

S

R

All isolates => % R = 11/17 = 65%

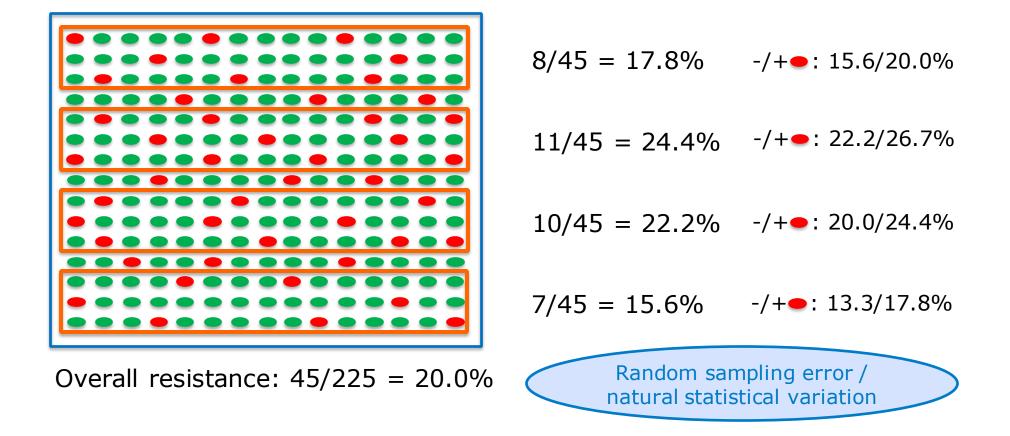
De-duplication

One isolate per patient => %R= 2/6= 33%

| Patient | Number of isolates in dataset |
|---------|-------------------------------|
| 1 | 5 |
| 2 | 2 |
| 3 | 2 |
| 4 | 1 |
| 5 | 6 |
| 6 | 1 |
| Total | 17 |

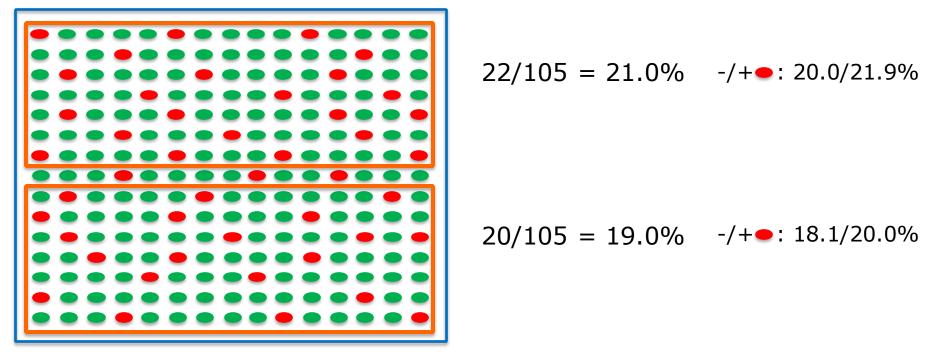


Sampling procedures – 4. sample size





Sampling procedures – 4. sample size



Overall resistance: 45/225 = 20.0%



Error and bias in national AMR surveillance

Surveillance system

- 1. Geographical region
- 2. Hospital types

Sampling procedures

- 1. Selection of patients
- 2. Sampling after treatment
- 3. Follow-up samples
- 4. Sample size

Laboratory procedures

- 1. AST guidelines
- 2. Measurement error
- 3. Selective testing



Laboratory procedures – 1. AST guidelines

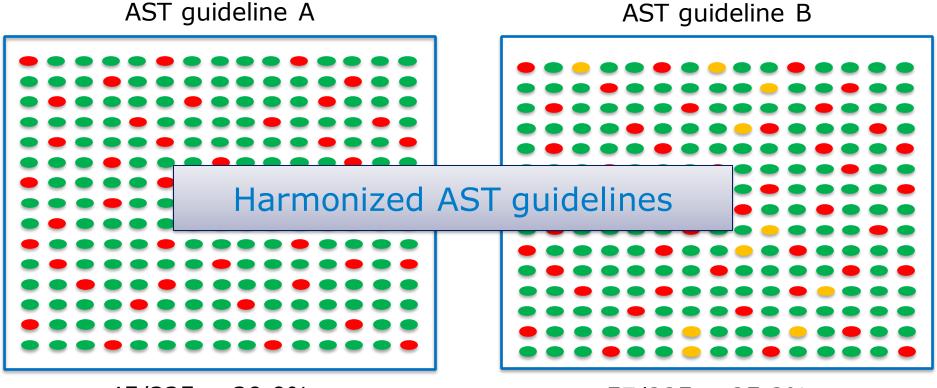
- EUCAST / CLSI / other guidelines
 - Breakpoints
 - Expert rules

| <i>E. coli</i> - Ceftazidime | | | | |
|------------------------------|--------|----------|--|--|
| Guideline | MIC Br | eakpoint | | |
| CLSI | S | ≤4 | | |
| | Ι | 8 | | |
| | R | ≥16 | | |
| EUCAST | S | ≤1 | | |
| | R | >4 | | |

\rightarrow different % SIR



Laboratory procedures – 1. AST guidelines

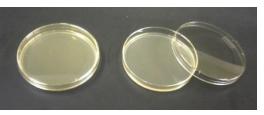


45/225 = 20.0%

57/225 = 25.3%



Laboratory procedures – 2. measurement error



Preparation and storage of agar plates



Inoculum

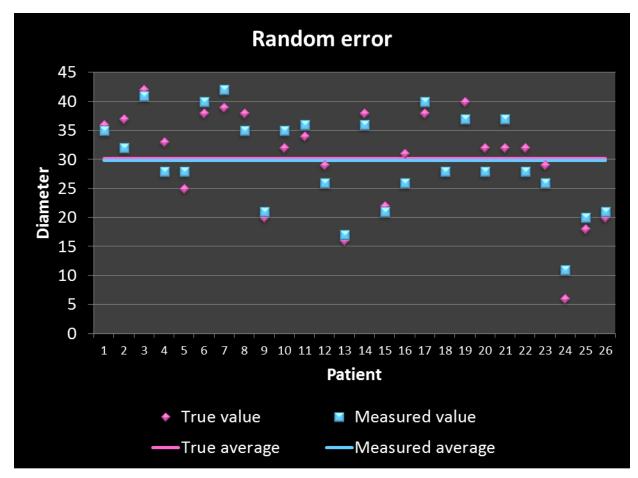


Uncalibrated automated system

Random vs. systematic measurement error

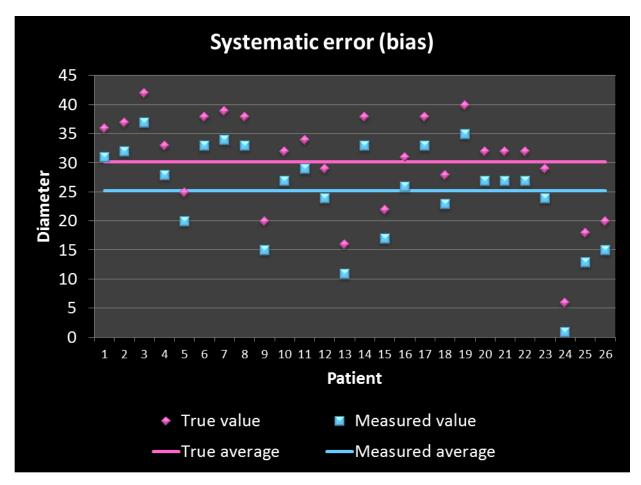


Laboratory procedures – 2. measurement error



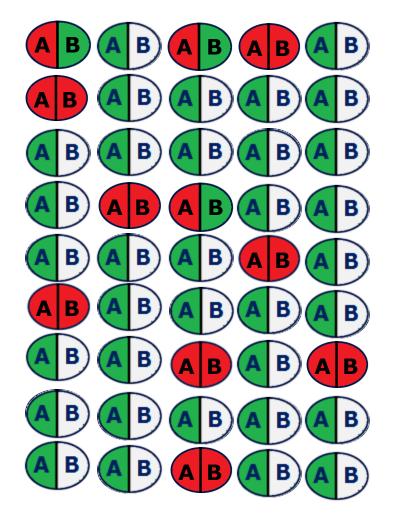


Laboratory procedures – 2. measurement error





Laboratory procedures – 3. selective testing



Testing all isolates against both drugs A: 11/45=24% B: 10/45=22% Sequential testing:

A: 11/45=24% B: 8/11=**73%**

- Check for and report on differences in numbers of isolates
- Exclude data for pathogen-antibiotic combination in lab, when tested for <50% of isolates of the pathogen



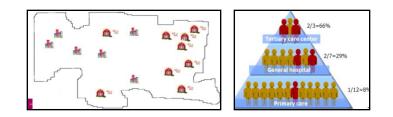
Most error and bias can not be corrected afterwards!

- 1. Prevent error and bias during data collection
 - Design of surveillance system
 - Case definitions for sampling
 - Standardize! Make consistent, uniform, comparable
- 2. Interpret results carefully
 - Understand how error and bias influence your data

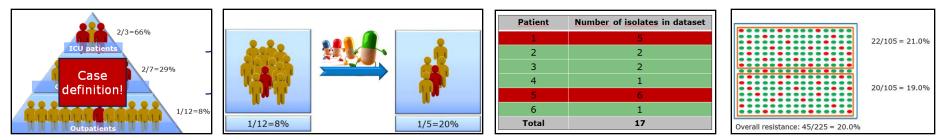


Interpretation and reporting of results – questions to ask yourself

- > How is the surveillance system designed?
 - Geographical representativeness
 - Selection of hospitals in surveillance



- > What sampling procedures were used?
 - Was a clear case definition formulated and adhered to?
 - Did sampling occur before treatment?
 - Are follow-up samples included? (exclude them from analyses!)
 - Is the sample size large enough for robust estimates?

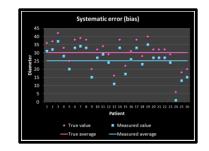


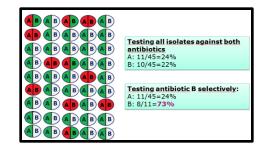


Interpretation and reporting of results – questions to ask yourself

- > Could laboratory procedures have caused bias / incomparable results?
 - Are harmonized AST guidelines and clinical breakpoints used?
 - Are exceptional phenotypes confirmed? (plausibility of laboratory results, i.e. indications for measurement error)
 - Are there indications for selective testing (are all isolates tested for all relevant antibiotics)?

| E. coli - Ceftazidime | | AST guideline B | | |
|-----------------------|--------|-----------------|--|----|
| Guideline | MIC Br | eakpoint | | :: |
| CLSI - | S | ≤4 | | |
| | I | 8 | | |
| | R | ≥16 | | |
| EUCAST - | S | ≤1 | | |
| | R | >4 | | |







Conclusion

- AMR surveillance data are extremely valuable in taking measures to prevent and control the spread of AMR
 - Awareness & advocacy
 - Infection prevention and control
- AMR surveillance data are subject to different types of error and bias
 - Magnitude unclear
 - Use of results for empiric therapy guidelines: with great caution!!!!!!
 - Interpret data carefully and report on how potential bias influenced the data

Thank you

